

Calyciphyllines H–M, new *Daphniphyllum* alkaloids from *Daphniphyllum calycinum*

Shizuka Saito^a, Hiroko Yahata^a, Takaaki Kubota^a, Yutaro Obara^b, Norimichi Nakahata^b, Jun'ichi Kobayashi^{a,*}

^a Graduate School of Pharmaceutical Sciences, Hokkaido University, Sapporo 060-0812, Japan

^b Graduate School of Pharmaceutical Sciences, Tohoku University, Aoba, Aramaki, Aoba-ku, Sendai 980-8578, Japan

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Abstract

Six new *Daphniphyllum* alkaloids, calyciphyllines H–M (**1–6**), were isolated from the leaves and stems of *Daphniphyllum calycinum* (Daphniphyllaceae). The structures and relative stereochemistry of **1–6** were elucidated on the basis of spectroscopic data, and the absolute stereochemistry of **3** was assigned by PGME method.

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Keywords: *Daphniphyllum calycinum*; *Daphniphyllum* alkaloid; Calyciphyllines H–M

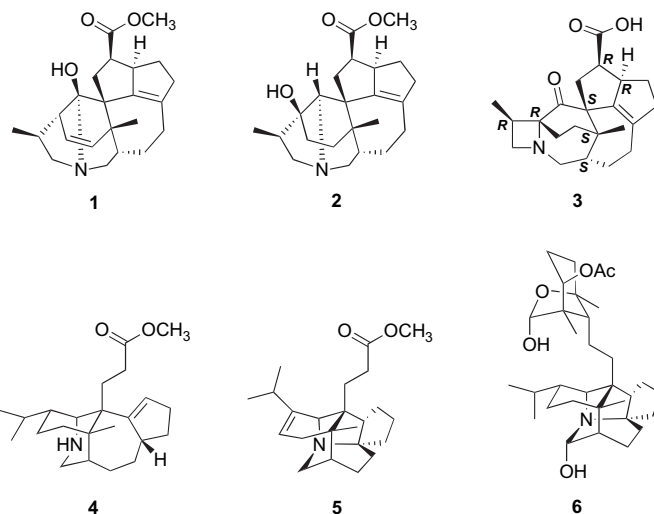
1. Introduction

Trees of the genus *Daphniphyllum* (Daphniphyllaceae) are known to elaborate structurally diverse group of alkaloids with unique polycyclic fused ring systems.^{1–6} These *Daphniphyllum* alkaloids have been attractive targets for biogenetic and synthetic studies.⁷ Recently, some novel alkaloids with unusual skeletons such as calyciphyllines C–G^{2a–d} have been isolated from *Daphniphyllum calycinum* at our laboratory. Further investigation of extracts of this plant resulted in the isolation of six new alkaloids, calyciphyllines H–M (**1–6**). In this paper we describe the isolation and structural elucidation of **1–6**.

2. Results and discussion

The leaves and stems of *D. calycinum* were extracted with MeOH separately, and each MeOH extract was partitioned between EtOAc and 3% tartaric acid. Water-soluble materials, which were adjusted to pH 10 with saturated Na₂CO₃, were extracted with CHCl₃ to give crude alkaloidal fractions. The crude alkaloidal materials prepared from the leaves were

subjected to an amino silica gel column (hexane/EtOAc and then CHCl₃/MeOH), in which a fraction eluted with hexane/EtOAc (6:4) was purified by a silica gel column (CHCl₃/MeOH) to give calyciphyllines H (**1**, 0.00007% yield), I (**2**, 0.00009%), K (**4**, 0.00003%), and L (**5**, 0.00018%). The crude alkaloidal materials prepared from the stems were separated by the same procedure as described above to yield calyciphyllines J (**3**, 0.00013%) and M (**6**, 0.00018%).



* Corresponding author. Tel.: +81 11 706 3239; fax: +81 11 706 4989.

E-mail address: jkobay@pharm.hokudai.ac.jp (J. Kobayashi).

Calyciphylline H (**1**) showed the pseudomolecular ion peak at m/z 370 ($M+H$)⁺ in the ESIMS, and the molecular formula, C₂₃H₃₁NO₃, was established by HRESIMS [m/z 370.2387, ($M+H$)⁺, Δ +0.5 mmu]. IR absorption at 1734 cm⁻¹ suggested the presence of the ester carbonyl functionality. Analyses of ¹H and ¹³C NMR data (Table 1) and the HMQC spectrum provided evidence that **1** possessed one tetrasubstituted olefin, one disubstituted olefin, one carbonyl, three sp³ quaternary carbons, five sp³ methines, seven sp³ methylenes, and three methyls. Among them, two methylenes (δ_C 54.7, δ_C 61.8) were ascribed to those bearing a nitrogen atom, while one quaternary carbon (δ_C 103.2) was assigned as an aminor carbon.

Three structural fragments, **a** (C-2 to C-4, C-2 to C-18, and C-18 to C-19 and C-20), **b** (C-6 to C-7 and C-12, and C-11 to C-12), and **c** (C-13 to C-17), were deduced from the ¹H–¹H COSY and TOCSY spectra as shown in Figure 1. HMBC correlations of H₂-7 to C-19 (δ_C 61.8) and H₂-19 to C-1 (δ_C 103.2) suggested that C-1, C-7, and C-19 were connected to each other through a nitrogen atom. Connections between C-4, C-6, and C-21 via C-5 were implied by HMBC cross-peaks for H-4 to C-5 (δ_C 41.8), and H₃-21 to C-4 (δ_C 138.0) and C-6 (δ_C 39.8). HMBC correlations for H-4 to C-8 (δ_C

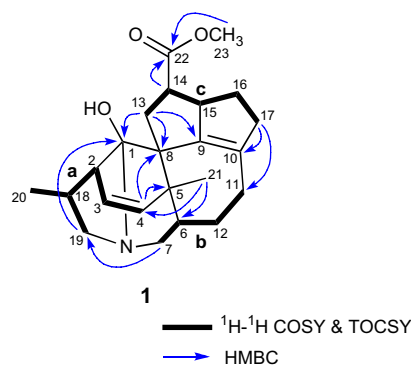


Figure 1. Selected 2D NMR correlations for calyciphylline H (**1**).

51.6), and H₂-13 to C-1, C-8, and C-9 (δ_C 141.9) indicated connectivities of C-1, C-5, C-9, and C-13 via C-8. The linkage of units **b** and **c** through C-10 was implied by HMBC cross-peaks for H₂-17 to C-10 (δ_C 137.4) and C-11 (δ_C 24.9). In addition, HMBC correlations for H₃-23 and H-14 to C-22 (δ_C 175.5) suggested that a methoxy group was attached to C-22. Thus, the gross structure of calyciphylline H was elucidated to be **1**.

Table 1
¹H and ¹³C NMR data of calyciphyllines H–J (**1–3**)

| | 1 ^a | | 2 ^a | | 3 ^a | |
|----|---------------------------|---------------------|-----------------------|------------|--------------------------|------------|
| | δ_H | δ_C | δ_H | δ_C | δ_H | δ_C |
| 1 | | 103.2 s | 3.89 (1H, br s) | 64.9 d | | 212.8 s |
| 2 | 2.99 (1H, m) | 46.4 d | | 76.8 s | | 75.2 s |
| 3 | 5.70 (1H, dd, 10.0, 3.5) | 125.0 d | 1.96 (1H, m) | 29.3 t | 2.32 (1H, m) | 23.0 t |
| | | | 1.80 (1H, m) | | 1.38 (1H, m) | |
| 4 | 5.49 (1H, dd, 10.0, 2.0) | 138.0 d | 1.96 (1H, m) | 35.7 t | 1.92 (1H, m) | 35.1 t |
| | | | 1.59 (1H, m) | | 1.82 (1H, m) | |
| 5 | | 41.8 s | | 34.4 s | | 39.4 s |
| 6 | 1.80 (1H, m) | 39.8 d | 2.13 (1H, m) | 41.3 d | 2.06 (1H, m) | 48.9 d |
| 7 | 3.46 (1H, dd, 13.5, 5.5) | 54.7 t | 3.76 (1H, m) | 55.8 t | 2.96 (1H, dd, 12.0, 4.6) | 54.7 t |
| | 3.31 (1H, m) | | 3.37 (1H, m) | | 2.58 (1H, m) | |
| 8 | | 51.6 s | | 44.9 s | | 59.9 s |
| 9 | | 141.9 s | | 141.9 s | | 142.3 s |
| 10 | | 137.4 s | | 136.0 s | | 133.6 s |
| 11 | 2.61 (1H, m) | 24.9 ^b t | 2.28 (1H, m) | 24.3 t | 2.06 (2H, m) | 25.4 t |
| | 2.14 (1H, m) | | 2.18 (1H, m) | | | |
| 12 | 2.14 (1H, m) | 25.1 ^b t | 2.18 (1H, m) | 27.5 t | 2.06 (1H, m) | 25.8 t |
| | 1.59 (1H, m) | | 1.53 (1H, m) | | 1.59 (1H, m) | |
| 13 | 2.61 (1H, m) | 39.3 t | 2.95 (1H, m) | 34.4 t | 2.74 (1H, dd, 13.7, 6.3) | 40.1 t |
| | 2.38 (1H, dd, 15.0, 10.0) | | 2.53 (1H, m) | | 1.92 (1H, m) | |
| 14 | 2.29 (1H, m) | 42.5 d | 2.91 (1H, m) | 41.5 d | 3.11 (1H, m) | 42.1 d |
| 15 | 3.76 (1H, m) | 58.1 d | 3.50 (1H, m) | 53.5 d | 4.01 (1H, m) | 56.4 d |
| 16 | 1.91 (1H, m) | 29.2 t | 1.87 (1H, m) | 27.8 t | 2.02 (1H, m) | 29.5 t |
| | 1.42 (1H, m) | | 1.24 (1H, m) | | 1.47 (1H, m) | |
| 17 | 2.61 (1H, m) | 43.2 t | 2.53 (1H, m) | 42.3 t | 2.61 (1H, m) | 42.5 t |
| | 2.31 (1H, dd, 15.0, 8.3) | | 2.28 (1H, m) | | 2.27 (1H, m) | |
| 18 | 3.16 (1H, m) | 32.4 d | 2.46 (1H, m) | 44.0 d | 3.05 (1H, m) | 25.9 d |
| 19 | 4.20 (1H, m) | 61.8 t | 4.13 (1H, m) | 61.1 t | 3.10 (2H, m) | 56.8 t |
| | 2.61 (1H, m) | | 2.53 (1H, m) | | | |
| 20 | 1.25 (3H, d, 7.3) | 18.3 q | 1.14 (3H, d, 6.9) | 12.4 q | 1.12 (3H, br d, 6.3) | 14.2 q |
| 21 | 1.38 (3H, s) | 22.6 q | 1.19 (3H, s) | 24.3 q | 1.19 (3H, s) | 26.3 q |
| 22 | | 175.5 s | | 175.6 s | | 178.9 s |
| 23 | 3.62 (3H, s) | 51.0 q | 3.63 (3H, s) | 51.3 q | | |

^a Measured in CDCl₃.

^b Assignments are interchangeable.

The relative stereochemistry of **1** was deduced from the NOESY spectrum as shown in Figure 2. NOESY correlations of H-3/H-4, H-3/H₃-20, H-4/H-6, H-4/H₃-21, and H-13a/H₃-21 indicated a pseudo boat conformation of cyclohexene ring (C-1 to C-5 and C-8) and a chair conformation of piperidine ring (C-1, N-1, and C-5 to C-8). Orientations of H-14 and H-15 were provided from NOESY correlations of H-13b/H-14 and H-14/H-15.

Calyciphylline I (**2**) showed the pseudomolecular ion peak at m/z 372 ($M+H$)⁺ in the ESIMS, and the molecular formula, C₂₃H₃₃NO₃, was established by HRESIMS [m/z 372.2552, ($M+H$)⁺, Δ +1.3 mmu]. The IR spectrum (1734 cm⁻¹) suggested the presence of ester carbonyl group. The ¹³C NMR data revealed 23 carbon signals due to one tetrasubstituted olefins, one carbonyl, three sp³ quaternary carbons, five sp³ methines, nine sp³ methylenes, and three methyls. Among them, one methine (δ_C 64.9) and two methylenes (δ_C 55.8, δ_C 61.1) were ascribed to those bearing a nitrogen atom.

The ¹H–¹H COSY and TOCSY spectra of **2** revealed connectivities of four partial structures, **a** (C-3 to C-4), **b** (C-6 to C-7 and C-12, and C-11 to C-12), **c** (C-13 to C17), and **d** (C-18 to C-19 and C-20) as shown in Figure 3. HMBC correlations of H₂-19 to C-1 (δ_C 64.9) and H₂-7 to C-19 (δ_C 61.1) suggested that C-1, C-7, and C-19 were connected to each other through a nitrogen atom. Connectivities of C-4, C-6, and C-21 via C-5 were implied by HMBC cross-peaks of H₃-21 to C-4 (δ_C 35.7), C-5 (δ_C 34.4), and C-6 (δ_C 41.3). HMBC correlations for H₂-13 to C-1, C-8 (δ_C 44.9), C-9 (δ_C 141.9) and H₃-21 to C-8 suggested that C-1, C-5, C-9, and C-13 were attached to C-8. Connections of C-11 and C-17 to C-9 through C-10 were implied by HMBC cross-peaks for H₂-16 to C-9, C-10 (δ_C 136.0), and H₂-12 to C-10.

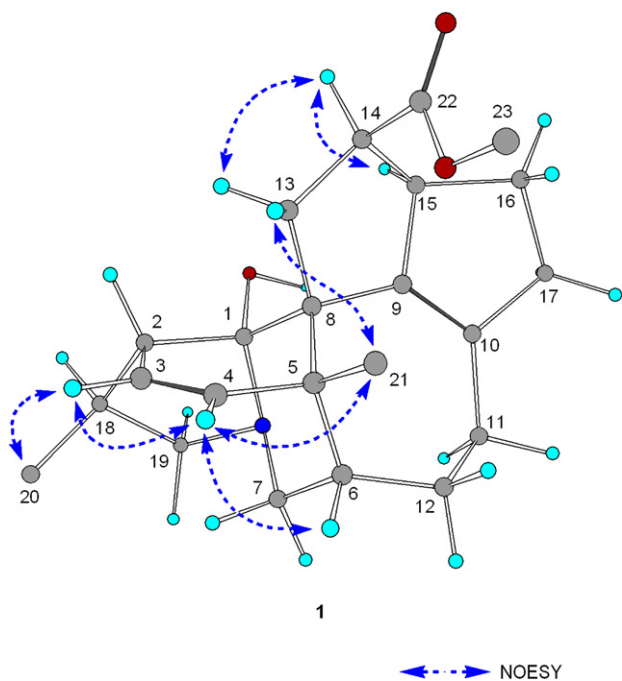


Figure 2. Selected NOESY correlations and relative stereochemistry of calyciphylline H (**1**) (hydrogen atoms of methyl groups were omitted).

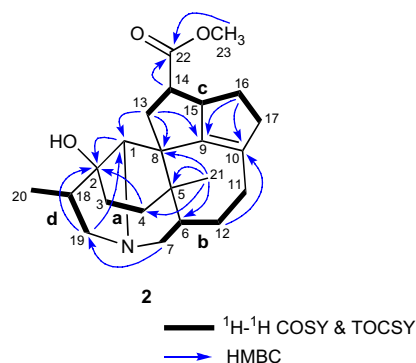


Figure 3. Selected 2D NMR correlations for calyciphylline I (**2**).

HMBC correlations of H-1, H₂-4, and H₂-19 to C-2 (δ_C 76.8) indicated the linkage of C-1 and C-3, C-18 via C-2. HMBC correlations for H₃-23 and H-14 to C-22 (δ_C 175.6) suggested that a methoxy group was attached to C-22. Thus, the gross structure of calyciphylline I was elucidated to be **2**.

The relative stereochemistry of **2** was elucidated from NOESY correlations as shown in Figure 4. Chair conformations of a cyclohexane ring (C-1 to C-5 and C-8) and a piperidine ring (C-1, N-1, and C-5 to C-8) were deduced from NOESY correlations of H₂-3/H₃-20, H-4b/H-6, H-6/H-7b, H-6/H₃-21, H-7b/H-19b, and H-19b/H₃-20. NOESY correlations of H-1/H-15, H-13a/H-14, and H-14/H-15 indicated α -orientations of H-14 and H-15.

Calyciphylline J (**3**) was obtained as a colorless amorphous solid, and the molecular formula was established as C₂₂H₂₉NO₃ by HRESIMS [m/z 356.2237, ($M+H$)⁺, Δ +1.1 mmu]. ¹H and ¹³C NMR data of **3** (Table 1) were similar to those of calyciphylline C,^{2d} except for the absence of a methoxy group. Detailed analyses of 2D NMR data indicated that **3** was the free acid form of calyciphylline C (Fig. 5).

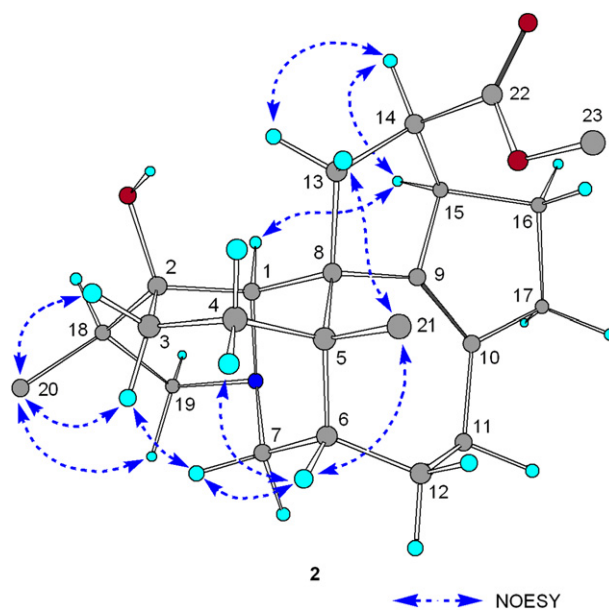


Figure 4. Selected NOESY correlations and relative stereochemistry of calyciphylline I (**2**) (hydrogen atoms of methyl groups were omitted).

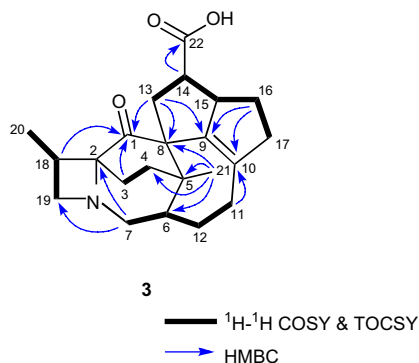


Figure 5. Selected 2D NMR correlations for calyciphylline J (**3**).

The relative stereochemistry of **3** was elucidated to be the same as that of calyciphylline C on the basis of NOESY data (Fig. 6).

To elucidate the absolute configuration at C-14, **3** was converted into its (*S*)- and (*R*)-phenylglycine methyl ester (PGME) amides of the carboxy group at C-14. The $\Delta\delta$ [δ (*S*-PGME amide)– δ (*R*-PGME amide)] values obtained from the ^1H NMR spectra of the PGME amides suggested that the absolute configuration at C-14 in **3** was *R* (Fig. 7).⁸ Thus, the absolute stereochemistry of **3** was elucidated as shown in Figure 7.

Calyciphylline J (**3**) was treated with trimethylsilyl diazomethane to afford calyciphylline C, whose spectral data and $[\alpha]_{\text{D}}^{20}$ value $\{[\alpha]_{\text{D}}^{20} -78.7$ (*c* 0.1, MeOH) $\}$ were coincident with those of natural calyciphylline C.^{2d,9} Thus, the absolute stereochemistry of calyciphylline C was concluded to be the same as **3**.

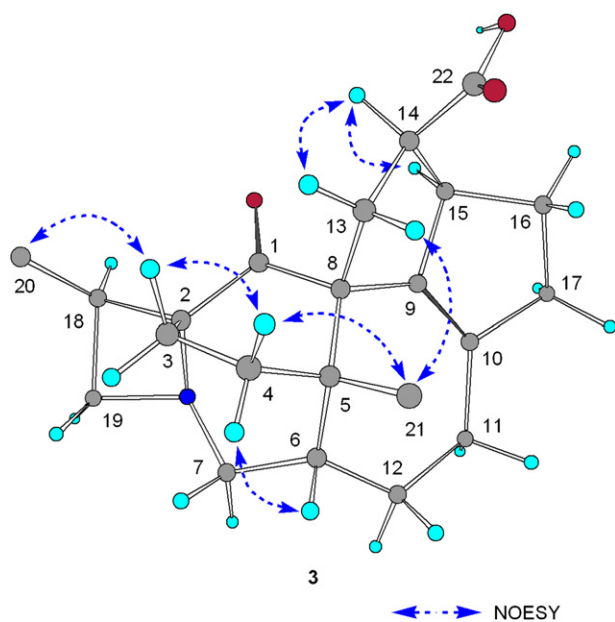


Figure 6. Selected NOESY correlations and relative stereochemistry of calyciphylline J (**3**) (hydrogen atoms of methyl groups were omitted).

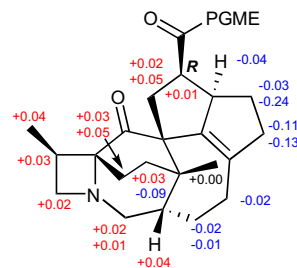


Figure 7. $\Delta\delta$ values [$\Delta\delta$ (in ppm)] $\delta_S - \delta_R$ obtained for the (*S*)- and (*R*)-PGME amides of calyciphylline J (**3**).

Calyciphylline K (**4**) showed the pseudomolecular ion peak at m/z 360 ($\text{M}+\text{H}$)⁺ in the ESIMS, and the molecular formula, $\text{C}_{23}\text{H}_{37}\text{NO}_2$, was established by HRESIMS [m/z 360.2905, ($\text{M}+\text{H}$)⁺, Δ +0.2 mmu]. The IR absorption at 1735 cm^{-1} suggested the presence of ester carbonyl functionality. The ^{13}C NMR (Table 2) spectrum of **4** gave signals due to one trisubstituted olefin, one carbonyl, two sp^3 quaternary carbons, five sp^3 methines, nine sp^3 methylenes, and four methyls, implying that the structure of **4** was similar to that of daphnezomine L.¹⁰

The chemical shifts of C-1 (δ_{C} 60.9) and C-7 (δ_{C} 47.5) in **4** suggested that these carbons were attached to a nitrogen atom and the connectivity of C-1 and C-7 via a nitrogen was implied by HMBC cross-peaks of H_2 -7 to C-1. The ^1H – ^1H COSY and TOCSY spectra of **4** revealed connectivities of three partial structures, **a** (C-1 to C-4, C-2 to C-18, and C-18 to C-19 and C-20), **b** (C-6 to C-7 and C-12, C-10 to C-12 and C-17, and C-15 to C-17), and **c** (C-13 to C-14), which were connected to each other on the basis of HMBC correlations as shown in Figure 8. HMBC correlations for H_3 -23 and H-14 to C-22 (δ_{C} 175.0) suggested that a methoxy group was attached to C-22. Thus, the gross structure of calyciphylline K (**4**) was elucidated to be *N*,7-dihydro daphnezomine L methyl ester.

The relative stereochemistry of **4** was elucidated from NOESY correlations as shown in Figure 9. Chair forms of a cyclohexane ring (C-1 to C-5 and C-8) and a piperidine ring (C-1, N-1, and C-5 to C-8), and a β -orientation of H-10 were suggested by NOESY cross-peaks of H-3b/H-7a and H-6/H₃-21 and H-10/H₃-21. An α -orientation of H-15 was suggested by NOESY cross-peaks of H-1/H-15 and H-1/H₃-20.

Calyciphylline L (**5**) showed the pseudomolecular ion peak at m/z 358 ($\text{M}+\text{H}$)⁺ in the ESIMS, and the molecular formula, $\text{C}_{23}\text{H}_{35}\text{NO}_2$, was established by HRESIMS [m/z 358.2740, ($\text{M}+\text{H}$)⁺, Δ –0.6 mmu]. The IR absorption at 1734 cm^{-1} suggested the presence of ester carbonyl functionality. The ^{13}C NMR data revealed 23 carbon signals due to one trisubstituted olefin, one ester carbonyl, three sp^3 quaternary carbons, four sp^3 methines, nine sp^3 methylenes, and four methyls. The chemical shifts of ^1H and ^{13}C NMR data (Table 2) suggested that the structure of **5** was close to that of methyl homodaphniphyllate.¹¹

The ^1H – ^1H COSY and TOCSY spectra of **5** revealed connectivities of five structural fragments, **a** (C-3 to C-4), **b** (C-6 to C-7 and C-12, and C-11 to C-12), **c** (C-9 to C-15, and C-15 to C-17), **d** (C-13 to C-14), and **e** (C-18 to C-19 and C-20) as

Table 2
 ^1H and ^{13}C NMR data of calyciphyllines K–M (4–6)

| | 4 ^a | | 5 ^b | | 6 ^a | |
|----|--------------------------|---------------------|-----------------------|---------------------|----------------------|---------------------|
| | δ_{H} | δ_{C} | δ_{H} | δ_{C} | δ_{H} | δ_{C} |
| 1 | 2.73 (1H, d, 3.5) | 60.9 d | 3.91 (1H, br s) | 66.9 d | 3.44 (1H, m) | 65.6 d |
| 2 | 1.40 (1H, m) | 41.0 d | | 136.0 s | 1.44 (1H, m) | 38.2 d |
| 3 | 1.80 (1H, m) | 27.1 t | 6.31 (1H, m) | 135.4 d | 1.87 (1H, m) | 26.0 t |
| | 1.55 (1H, m) | | | | 1.44 (1H, m) | |
| 4 | 1.96 (1H, m) | 41.0 t | 2.48 (1H, m) | 43.5 t | 1.98 (1H, m) | 36.1 t |
| | 1.40 (1H, m) | | 2.18 (1H, m) | | 1.65 (1H, m) | |
| 5 | | 37.9 s | | 39.5 s | | 38.2 s |
| 6 | 1.49 (1H, m) | 44.1 d | 1.76 (1H, m) | 42.3 d | 1.88 (1H, m) | 45.5 d |
| 7 | 3.44 (1H, dd, 14.7, 7.6) | 47.5 t | 3.58 (1H, br d, 12.6) | 44.6 t | 5.74 (1H, br d, 3.4) | 81.2 d |
| | 2.60 (1H, br d, 14.7) | | 3.14 (1H, br d, 12.6) | | | |
| 8 | | 46.6 s | | 49.1 s | | 47.1 s |
| 9 | | 155.5 s | 2.70 (1H, m) | 51.9 d | 2.48 (1H, m) | 51.5 d |
| 10 | 3.02 (1H, m) | 48.2 d | | 80.3 s | | 76.7 s |
| 11 | 1.72 (1H, m) | 34.7 t | 2.07 (1H, m) | 28.9 t | 2.76 (1H, m) | 28.7 t |
| | 1.43 (1H, m) | | 1.83 (1H, m) | | 1.64 (1H, m) | |
| 12 | 1.90 (1H, m) | 32.9 t | 2.07 (1H, m) | 22.1 t | 2.04 (1H, m) | 16.4 t |
| | 1.39 (1H, m) | | 1.83 (1H, m) | | 1.80 (1H, m) | |
| 13 | 2.13 (2H, m) | 31.7 t | 1.82 (1H, m) | 27.1 t | 2.10 (1H, m) | 22.9 t |
| | | | 1.54 (1H, m) | | 1.39 (1H, m) | |
| 14 | 2.32 (1H, m) | 31.7 t | 2.63 (1H, m) | 32.6 t | 1.94 (1H, m) | 32.3 t |
| | 1.63 (1H, m) | | 2.52 (1H, m) | | 1.00 (1H, m) | |
| 15 | 5.49 (1H, br s) | 126.5 d | 2.01 (1H, m) | 31.0 t | 1.96 (1H, m) | 29.4 t |
| | | | 1.75 (1H, m) | | 1.52 (1H, m) | |
| 16 | 2.37 (1H, m) | 29.8 t | 2.03 (1H, m) | 26.9 t | 1.90 (1H, m) | 25.1 t |
| | 2.18 (1H, m) | | 1.62 (1H, m) | | 1.52 (1H, m) | |
| 17 | 2.15 (1H, m) | 32.6 t | 2.12 (1H, m) | 40.9 t | 2.16 (1H, m) | 39.4 t |
| | 1.49 (1H, m) | | 1.92 (1H, m) | | 1.90 (1H, m) | |
| 18 | 1.67 (1H, m) | 31.1 d | 2.32 (1H, m) | 34.9 d | 1.70 (1H, m) | 29.8 d |
| 19 | 0.93 (3H, d, 6.9) | 20.9 q | 1.16 (3H, d, 6.4) | 22.2 q | 0.94 (3H, d, 6.3) | 20.9 q |
| 20 | 0.92 (3H, d, 6.9) | 20.9 q | 1.14 (3H, d, 6.4) | 23.1 q | 1.00 (3H, d, 6.3) | 21.7 q |
| 21 | 1.08 (3H, s) | 27.5 q | 1.17 (3H, s) | 24.3 q | 1.17 (3H, s) | 26.0 q |
| 22 | | 175.0 s | | 176.1 s | 1.59 (1H, m) | 52.0 d |
| 23 | 3.66 (3H, s) | 51.5 q | 3.69 (3H, s) | 53.1 q | | 50.0 s |
| 24 | | | | | 1.05 (3H, s) | 17.0 q |
| 25 | | | | | 4.88 (1H, br s) | 99.2 d |
| 26 | | | | | 4.79 (1H, br d, 5.2) | 73.6 d |
| 27 | | | | | 1.95 (1H, m) | 25.6 t |
| | | | | | 1.62 (1H, m) | |
| 28 | | | | | 1.53 (1H, m) | 27.5 t |
| | | | | | 1.35 (1H, m) | |
| 29 | | | | | | 84.2 s |
| 30 | | | | | 1.33 (3H, s) | 26.0 q |
| 31 | | | | | | 169.9 s |
| 32 | | | | | 2.09 (3H, s) | 21.6 q |

^a Measured in CDCl_3 .

^b Measured in CD_3OD .

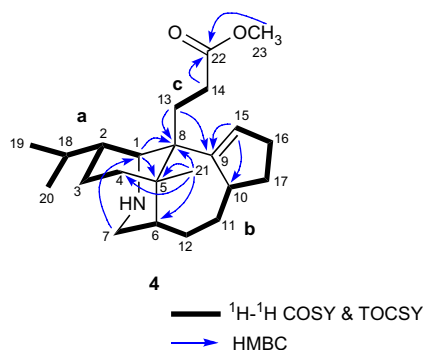


Figure 8. Selected 2D NMR correlations for calyciphylline K (4).

shown in Figure 10. Connections between C-1, C-3, and C-18 via C-2 were implied by HMBC cross-peaks for H-3 to C-1 (δ_{C} 66.9) and C-18 (δ_{C} 34.9), and H₃-19 to C-2 (δ_{C} 136.0). The connectivities of five partial structures a–e were revealed on the basis of HMBC correlations as shown in Figure 10. In addition, HMBC correlations for H₃-23 and H₂-14 to C-22 (δ_{C} 176.1) suggested that a methoxy group was attached to C-22. Thus, the gross structure of calyciphylline L (5) was elucidated to be 2,3-dehydro methyl homodaphniphyllate.

The relative stereochemistry of 5 was elucidated from NOESY correlations as shown in Figure 11. A pseudo boat conformation of a cyclohexene ring (C-1 to C-5 and C-8),

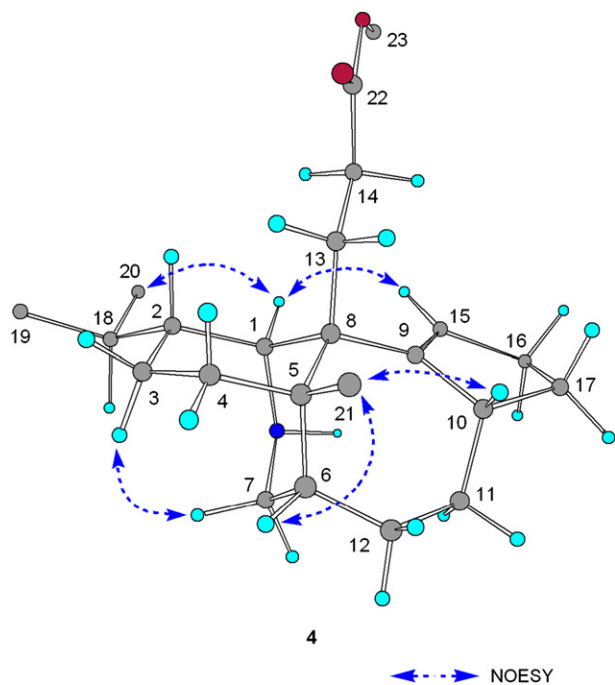


Figure 9. Selected NOESY correlations and relative stereochemistry of calyciphylline K (**4**) (hydrogen atoms of methyl groups were omitted).

a chair conformation of a piperidine ring (C-1, N-1, and C-5 to C-8), and a β -orientation of H-9 were deduced from NOESY cross-peaks of H-4b/H-6, H-4b/H-7a, H-6/H₃-21, and H-9/H₃-21.

The molecular formula of calyciphylline M (**6**) was determined to be C₃₂H₅₁NO₅ by HRESIMS [m/z 530.3850, (M+H)⁺, Δ +0.5 mmu]. The IR spectrum suggested the presence of hydroxyl (3300 cm⁻¹) and ester carbonyl (1731 cm⁻¹) groups.

¹H and ¹³C NMR, and the HMQC spectra revealed that **6** possessed 23 carbons due to one ester carbonyl, five sp³ quaternary carbons, nine sp³ methines, 11 sp³ methylenes, and six methyls. The chemical shifts of ¹H and ¹³C NMR data (Table 2) implying that **6** had the same fused-pentacyclic backbone skeleton as that of daphnimacropine¹² with the side chain

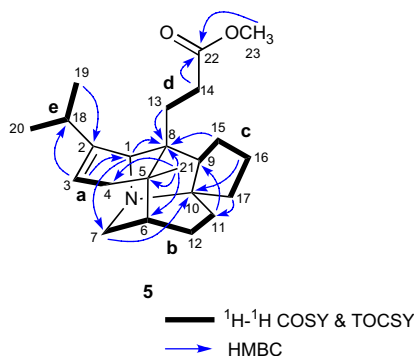


Figure 10. Selected 2D NMR correlations for calyciphylline L (**5**).

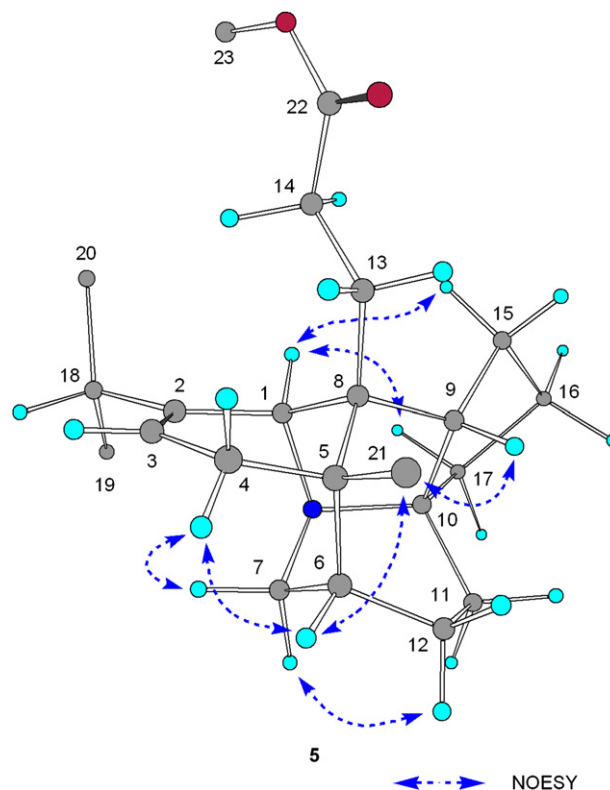


Figure 11. Selected NOESY correlations and relative stereochemistry of calyciphylline L (**5**) (hydrogen atoms of methyl groups were omitted).

consistent with that of daphmacropodine.¹³ Detailed analyses of 2D NMR data revealed that **6** was the 7-hydroxy form of daphmacropodine (Figs. 12 and 13).

Effects of calyciphyllines H–M (**1–6**) on neurotrophic factor biosynthesis in 1321N1 human astrocytoma cells were examined by a semiquantitative RT-PCR method.^{14,15} Among compounds **1–6**, it was found that the mRNA expressions for NGF were significantly enhanced by **4**.

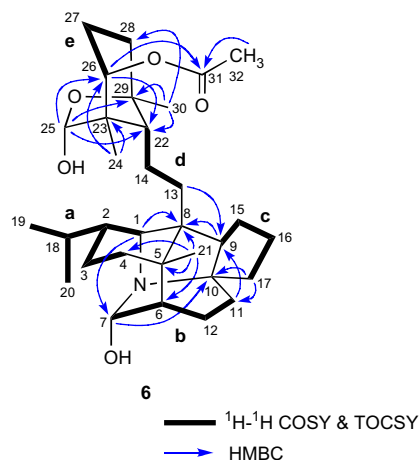


Figure 12. Selected 2D NMR correlations for calyciphylline M (**6**).

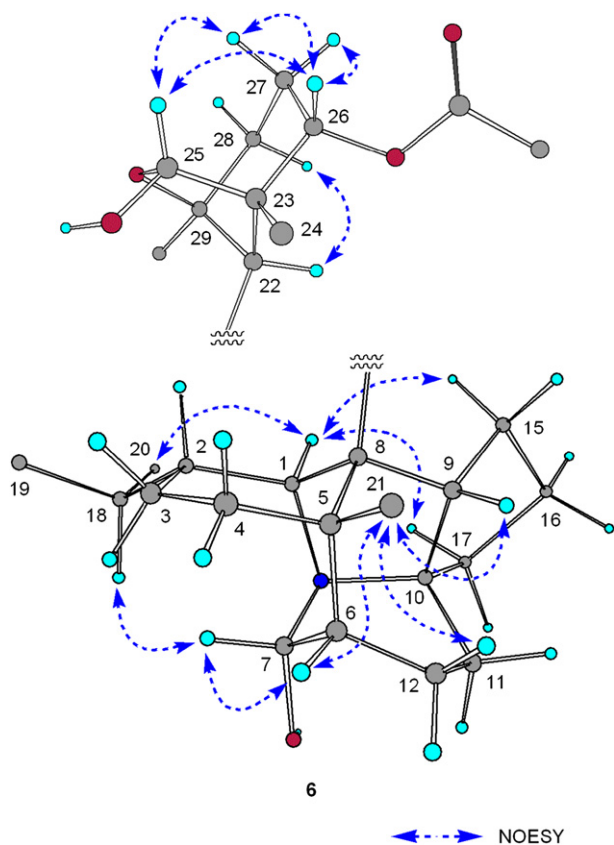


Figure 13. Selected NOESY correlations and partial relative stereochemistry of calyciphylline M (**6**) (hydrogen atoms of methyl groups were omitted).

3. Experimental

3.1. General experimental procedures

Optical rotation was recorded on a JASCO P-1030 polarimeter. IR and UV spectra were recorded on JASCO FT/IR-230 and Shimadzu UV-1600PC spectrophotometers, respectively. ^1H and ^{13}C NMR spectra were recorded on a Bruker AMX-600 and a JEOL ECA-500 spectrometers. The 7.26 and 77.0 ppm resonances of residual CDCl_3 and the 3.35 and 49.8 ppm resonances of residual CD_3OD were used as internal references for ^1H and ^{13}C NMR spectra, respectively. ESI mass spectra were obtained on a JEOL JMS-700TZ spectrometer.

3.2. Isolation

The leaves and stems of *D. calycinum* were extracted with MeOH, and the MeOH extract was partitioned between EtOAc and 3% tartaric acid. Water-soluble materials, which are adjusted to pH 10 with saturated Na_2CO_3 , were extracted with CHCl_3 to give crude alkaloidal fractions. The crude alkaloidal fraction prepared from the leaves was subjected to an amino silica gel column (hexane/EtOAc, 1:0→4:6, and then $\text{CHCl}_3/\text{MeOH}$, 1:0→0:1), in which a fraction eluted with hexane/EtOAc (6:4) was purified by a silica gel column ($\text{CHCl}_3/\text{MeOH}$, 1:0→0:1) to give calyciphyllines H (**1**, 0.00007%

yield), I (**2**, 0.00009%), K (**4**, 0.00003%), and L (**5**, 0.00018%). The crude alkaloidal fraction prepared from the stems was separated by the same procedure as described above to yield calyciphyllines J (**3**, 0.00013%) and M (**6**, 0.00018%).

3.2.1. Calyciphylline H (**1**)

Colorless amorphous solid; $[\alpha]_{\text{D}}^{22} +36.6$ (*c* 0.5, CHCl_3); IR (neat) ν_{max} 3649, 2928, 1734, 1669, 1457, and 1200 cm^{-1} ; ^1H and ^{13}C NMR, see Table 1; ESIMS m/z 370 ($\text{M}+\text{H}$) $^+$; HRESIMS m/z 370.2387 ($\text{M}+\text{H}$; calcd for $\text{C}_{23}\text{H}_{32}\text{NO}_3$, 370.2382).

3.2.2. Calyciphylline I (**2**)

Colorless amorphous solid; $[\alpha]_{\text{D}}^{22} -9.2$ (*c* 0.5, CHCl_3); IR (neat) ν_{max} 3394, 2964, 1734, 1673, 1457, 1200, and 1134 cm^{-1} ; ^1H and ^{13}C NMR data, see Table 1; ESIMS m/z 372 ($\text{M}+\text{H}$) $^+$; HRESIMS m/z 372.2552 ($\text{M}+\text{H}$; calcd for $\text{C}_{23}\text{H}_{34}\text{NO}_3$, 372.2539).

3.2.3. Calyciphylline J (**3**)

Colorless amorphous solid; $[\alpha]_{\text{D}}^{22} -45.0$ (*c* 1.0, CHCl_3); IR (neat) ν_{max} 3420, 2931, 1735, 1705, 1456, 1384, and 1194 cm^{-1} ; ^1H and ^{13}C NMR data, see Table 1; ESIMS m/z 356 ($\text{M}+\text{H}$) $^+$; HRESIMS m/z 356.2237 ($\text{M}+\text{H}$; calcd for $\text{C}_{22}\text{H}_{30}\text{NO}_3$, 356.2226).

3.2.4. (*R*)- and (*S*)-PGME amides of calyciphylline J (**3**)

To an ice cooled DMF solution (500 μL) of **3** (0.4 mg) and (*S*)-PGME (0.5 mg) were added PyBOP (1.0 mg), HOBt (0.5 mg), and *N*-methylmorpholine (25 μL), and stirring was continued at rt for 27 h. After addition of 5% HCl (1 mL), the mixture was extracted with EtOAc (2.5 mL). The extract was washed with saturated NaHCO_3 aq (1 mL) and brine (1 mL), and then concentrated in vacuo to afford the (*S*)-PGME amide of **3** (0.6 mg). The (*R*)-PGME amide of **3** was prepared according to the same procedure as described above.

3.2.5. (*R*)-PGME amide of calyciphylline J (**3**)

^1H NMR (600 MHz in CDCl_3) δ 7.33 (5H, m, phenyl), 6.39 (1H, d, 7.2, NH), 5.57 (1H, d, 7.2, CHNH), 3.96 (1H, m, H-15), 3.72 (3H, s, OCH_3), 3.06 (2H, m, H₂-19), 3.01 (1H, m, H-18), 2.94 (1H, m, H-14), 2.80 (2H, m, H-7, H-13), 2.59 (1H, m, H-17), 2.53 (1H, m, H-7), 2.27 (1H, dd, 15.0, 8.5, H-17), 2.23 (1H, m, H-3), 2.00 (3H, m, H₂-11, H-12), 1.98 (1H, m, H-4), 1.95 (1H, m, H-16), 1.90 (1H, m, H-6), 1.82 (1H, m, H-13), 1.73 (1H, m, H-4), 1.66 (1H, m, H-12), 1.45 (1H, m, H-16), 1.33 (1H, m, H-3), 1.19 (3H, s, H₃-21), 1.03 (3H, d, 6.2, H₃-20); ESIMS m/z 503 ($\text{M}+\text{H}$) $^+$; HRESIMS m/z 503.29018 ($\text{M}+\text{H}$; calcd for $\text{C}_{31}\text{H}_{39}\text{N}_2\text{O}_4$, 503.29098).

3.2.6. (*S*)-PGME amide of calyciphylline J (**3**)

^1H NMR (600 MHz in CDCl_3) δ 7.34 (5H, m, phenyl group), 6.35 (1H, d, 6.5, NH), 5.54 (1H, d, 7.2, CHNH), 3.92 (1H, m, H-15), 3.71 (3H, s, OCH_3), 3.08 (2H, m, H₂-19), 3.04 (1H, m, H-18), 2.95 (1H, m, H-14), 2.82 (2H, m, H-7, H-13), 2.54 (1H, m, H-7), 2.48 (1H, m, H-17), 2.26 (1H, m, H-3), 2.14 (1H, dd, 14.9, 8.4, H-17), 1.98 (3H, m,

H₂-11, H-12), 1.94 (1H, m, H-6), 1.89 (1H, m, H-4), 1.87 (1H, m, H-13), 1.76 (1H, m, H-4), 1.65 (2H, m, H-12, H-16), 1.38 (1H, m, H-3), 1.21 (1H, m, H-16), 1.19 (3H, s, H₃-21), 1.07 (3H, d, 6.5, H₃-20); ESIMS *m/z* 503 (M+H)⁺; HRESIMS *m/z* 503.28919 (M+H; calcd for C₃₁H₃₉N₂O₄, 503.29098).

3.2.7. Chemical conversion of calyciphylline J (3) to calyciphylline C

To a MeOH solution (300 μL) of **3** (0.4 mg) was added two drops of trimethylsilyl diazomethane (2.0 M solution in diethyl ether), and stirring was continued at rt for 2 h. The mixture was concentrated in vacuo, and the residue was purified by a silica gel column (CHCl₃/MeOH, 1:0→0:1) to afford calyciphylline C (0.2 mg).

3.2.8. Calyciphylline K (4)

Colorless amorphous solid; $[\alpha]_{\text{D}}^{19} -38.1$ (*c* 1.0, CHCl₃); IR (neat) ν_{max} 2923, 1735, 1543, 1508, 1457, and 1169 cm⁻¹; ¹H and ¹³C NMR data, see Table 2; ESIMS *m/z* 360 (M+H)⁺; HRESIMS *m/z* 360.2905 (M+H; calcd for C₂₃H₃₈NO₂, 360.2903).

3.2.9. Calyciphylline L (5)

Colorless amorphous solid; $[\alpha]_{\text{D}}^{22} -3.5$ (*c* 0.5, CHCl₃); IR (neat) ν_{max} 2961, 1734, 1672, 1458, 1323, 1199, and 1130 cm⁻¹; ¹H and ¹³C NMR data, see Table 2; ESIMS *m/z* 358 (M+H)⁺; HRESIMS *m/z* 358.2740 (M+H; calcd for C₂₃H₃₆NO₂, 358.2746).

3.2.10. Calyciphylline M (6)

Colorless amorphous solid; $[\alpha]_{\text{D}}^{17} +1.1$ (*c* 1.0, CHCl₃); IR (neat) ν_{max} 3300, 2961, 1731, 1665, 1456, 1242, 1201, and 1136 cm⁻¹; ¹H and ¹³C NMR data, see Table 2; ESIMS *m/z* 530 (M+H)⁺; HRESIMS *m/z* 530.3850 (M+H; calcd for C₃₂H₅₂NO₅, 530.3845).

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References and notes

- For a review of *Daphniphyllum* alkaloids see: Kobayashi, J.; Morita, H. *The Alkaloid*; Cordell, G. A., Ed.; Academic: New York, NY, 2003; Vol. 60, pp 165–205 and references therein.
- (a) Saito, S.; Kubota, T.; Zhang, H.; Kobayashi, J. *Tetrahedron Lett.* **2007**, *48*, 5693–5695; (b) Saito, S.; Kubota, T.; Zhang, H.; Kobayashi, J. *Tetrahedron Lett.* **2007**, *48*, 3809–3812; (c) Saito, S.; Kubota, T.; Fukushima, E.; Kawabata, J.; Zhang, H.; Kobayashi, J. *Org. Lett.* **2007**, *9*, 1207–1209; (d) Saito, S.; Kubota, T.; Fukushima, E.; Kawabata, J.; Zhang, H.; Kobayashi, J. *Tetrahedron Lett.* **2007**, *48*, 1587–1589; (e) Morita, H.; Ishioka, N.; Takatsu, H.; Iizuka, T.; Kobayashi, J. *J. Nat. Prod.* **2006**, *69*, 418–420; (f) Kubota, T.; Matsuno, Y.; Morita, H.; Shinzato, T.; Sekiguchi, M.; Kobayashi, J. *Tetrahedron* **2006**, *62*, 4743–4748; (g) Morita, H.; Ishioka, N.; Takatsu, H.; Shinzato, T.; Obara, Y.; Nakahata, N.; Kobayashi, J. *Org. Lett.* **2005**, *7*, 459–462; (h) Takatsu, H.; Morita, H.; Shen, Y. C.; Kobayashi, J. *Tetrahedron* **2004**, *60*, 6279–6284; (i) Morita, H.; Takatsu, H.; Shen, Y. C.; Kobayashi, J. *Tetrahedron Lett.* **2004**, *45*, 901–904; (j) Morita, H.; Kobayashi, J. *Org. Lett.* **2003**, *5*, 2895–2898; (k) Kobayashi, J.; Takatsu, H.; Shen, Y. C.; Morita, H. *Org. Lett.* **2003**, *5*, 1733–1736; (l) Morita, H.; Takatsu, H.; Kobayashi, J. *Tetrahedron* **2003**, *59*, 3575–3579.
- Bitar, H. E.; Nguyen, V. H.; Gramain, A.; Sévenet, T.; Bodo, B. *Tetrahedron Lett.* **2004**, *45*, 515–518.
- Li, Z.-Y.; Chen, P.; Xu, H.-G.; Yang, Y.-M.; Peng, S.-Y.; Zhao, Z.-Z.; Guo, Y.-W. *Org. Lett.* **2007**, *9*, 477–480.
- Di, Y. T.; He, H. P.; Lu, C. S.; Tian, J. M.; Mu, S. Z.; Li, S. L.; Gao, S.; Hao, X. J. *J. Nat. Prod.* **2006**, *69*, 1745–1748.
- Fan, C. Q.; Yin, S.; Xue, J. J.; Yue, J. M. *Tetrahedron* **2007**, *63*, 115–119.
- (a) Wallace, G. A.; Heathcock, C. H. *J. Org. Chem.* **2001**, *66*, 450–454; (b) Heathcock, C. H. *Proc. Natl. Acad. Sci. U.S.A.* **1996**, *93*, 14323–14327; (c) Heathcock, C. H.; Joe, D. *J. Org. Chem.* **1995**, *60*, 1131–1142; (d) Heathcock, C. H.; Kath, J. C.; Ruggeri, R. B. *J. Org. Chem.* **1995**, *60*, 1120–1130; (e) Heathcock, C. H. *Angew. Chem.* **1992**, *104*, 675–691; (f) Heathcock, C. H. *Angew. Chem., Int. Ed. Engl.* **1992**, *31*, 665–681 and references cited therein.
- Nagai, Y.; Kusumi, T. *Tetrahedron Lett.* **1995**, *36*, 1853–1856.
- Repurified natural calyciphylline C showed a relatively larger $[\alpha]_{\text{D}}$ value $\{[\alpha]_{\text{D}}^{20} -79.6$ (*c* 0.1, MeOH) $\}$ as compared with the value previously reported for calyciphylline C.^{2d}
- Morita, H.; Kobayashi, J. *Tetrahedron* **2002**, *58*, 6637–6641.
- Toda, M.; Niwa, H.; Irikawa, H.; Hirata, Y.; Yamamura, S. *Tetrahedron* **1974**, *30*, 2683–2688.
- Kamijo, N.; Nakano, T.; Terao, Y.; Osaki, K. *Tetrahedron Lett.* **1966**, *25*, 2889–2892.
- (a) Nakano, T.; Hasegawa, M.; Saeki, Y. *J. Org. Chem.* **1973**, *38*, 2404–2405; (b) Yang, S. P.; Yue, J. M. *Helv. Chim. Acta* **2006**, *89*, 2783–2788.
- Obara, Y.; Kobayashi, H.; Ohta, T.; Ohizumi, Y.; Nakahata, N. *Mol. Pharmacol.* **2001**, *59*, 1287–1297.
- Morita, H.; Ishiuchi, K.; Haganuma, A.; Hoshino, T.; Obara, Y.; Nakahata, N.; Kobayashi, J. *Tetrahedron* **2005**, *61*, 1955–1960.